# College of Forestry | Forest Ecosystems and Society

# Just rol with it: A transgene excision system for rapid transformation and gene-editing in plants

### INTRODUCTION

Plant genetic transformation is a useful tool for improving resistance to disease and abiotic stressors like drought and heat, but remains difficult or impossible in the large majority of plant species. Insertion of *Rhizobium rhizogenes* genes reliably produces transgenic hairy roots in many plant species, but generating transgenic shoots from hairy root tissue is challenging. In addition, the presence of the *rol* gene cassette in the Ri plasmid after shoot production leads to phenotypic abnormalities. We therefore developed a heat-inducible system for removal of undesired genes, including the *rol* gene cassette, using the Cre/lox recombinase system. The system appears to be working well in transgenic poplars.



### **VECTOR ASSEMBLY**



- To create a heat-shock-inducible Cre/lox recombinase system, we assembled a complex transformation vector between two recombinase recognition sites which undergo excision upon a heat shock treatment.
- Constructs were generated using the **Golden Gate method**, which allows rapid and seamless assembly of plant vectors, and further assembled using **GAANTRY**, a system for stably stacking genes in the Agrobacterium T-DNA.
- Gene editing via CRISPR/Cas9 aims to produce pollen and seed-free poplars with reduced stature

### **TRANSGENE EXCISION**



1. Transgene is excised as a non-chromosomal DNA fragment and a small "footprint" is left in the genome



2. Shoot development is induced and Cre recombinase initiates recombination between lox sites



electrophoresis

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Hairy root from a single plated explant

### Longer heat-shock (HS) duration improved shoot regeneration rates per explant (segments of hairy roots)





- excision via Cre recombinase
- indicate that excision was successful



An explant in full-color (left) and overlaid fluorescent signals (right)

Presence of the excision "footprint" verified by PCR and gel

Three transformation markers (one magenta pigment as shown below, and two fluorescent markers) allowed for easy identification of transgenic, excised tissue

 Hairy roots expressing the RUBY phenotype (magenta) are transgenic • Explants were exposed to three different heat shock treatments to induce

• Absence of fluorescent transformation markers GFP and DsRed (below)



No heat-shock



## **RESULTS SUMMARY**

- 36% were still transgenic or chimeric
- 14% were WT escapes

### **CURRENT RESEARCH**

We are testing the efficacy of the novel construct in multiple poplar genotypes and investigating the relative importance of its genetic components in isolation (WUS, ipt, Cre).

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The 14-day heat-shock (HS) resulted in highest rate of shoot regeneration

One representative plate from each heat shock treatment from below

### Regeneration rate among all explants

### **Heat Shock Duration**

• 48% of events had at least one propagatable, visually excised shoot, most had many more Of the shoots we screened at the DNA level: 50% were excised





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