## Toward Gene Editing in Hop:

Application of a Cutting-Edge Tool for Accelerating Breeding and Improving Traits

> Presented by: Chris Willig Oregon State University



## Genetic engineering vs. gene editing

Genetic engineering / transformation - method for delivering a "package" of genetic material into a plant to alter a trait

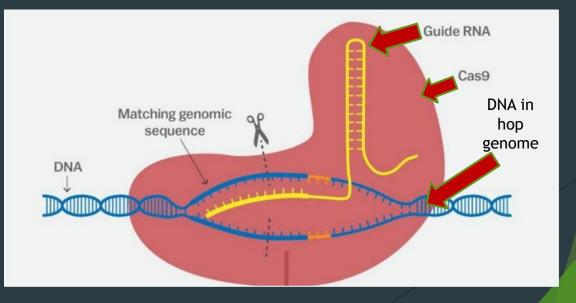
Gene editing is using the package to deliver "machinery" that then changes genes already inside the plant to alter a trait



### How CRISPR works at the molecular level

CRISPR machinery consists of a DNA cutting component (Cas protein) and a sequence recognition component (guide RNA)

CRISPR is a very customizable tool – to edit a new gene we need only to add a unique ~20-base sequence corresponding to guide RNA



## Why use biotech in hop?

Hop agriculture is facing threats due to a changing global climate

Extreme temperature waves

Periodic drought

Disease and pest outbreak





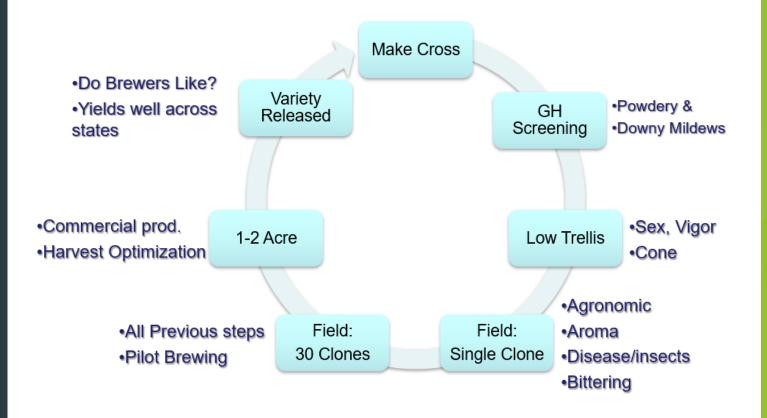
Can be addressed by breeding hop varieties with improved traits that offer some protection from these pressures

## The hop breeding cycle

Breeding may take 8-15 years between an initial cross and commercial release of a new cultivar

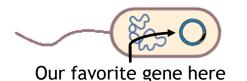
Deals with traits influenced by many genes

Very difficult to incorporate new traits while maintaining aroma profile of prized cultivars

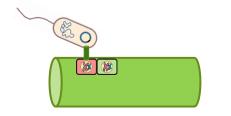


## Transforming plants with Agrobacterium

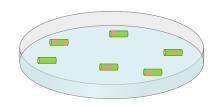
A selected gene sequence is inserted into an *Agrobacterium* cell



Agrobacterium delivers target gene to plant cells



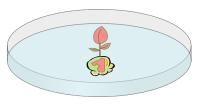
When plant tissue is initially plated, relatively few cells are modified with the selected gene



Transformed shoots can be rooted and grown into a plant that is now permanently modified



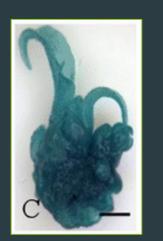
"Transformed" shoots are formed from populations of cells with the selected gene



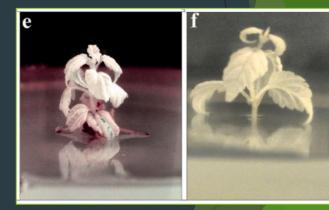
As callus forms, cells that have the gene become more abundant through selection

## Prior work in hop transformation

- A few labs have reported successful transformation of hop, all in European cultivars
- The first report of successful gene editing in hop was published by a Czech group in 2021
- This is encouraging, but methods need to be adapted to each different cultivar
- We were awarded a USDA-NIFA grant in 2021 to develop transformation/gene editing methods in US hop varieties



Transgenic shoot of 'Tettnanger' (Arias & Weber, 2013)



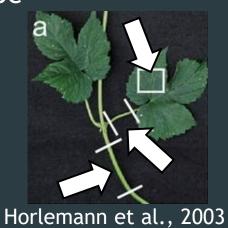
Gene edited hop plants for PDS gene (proof-ofconcept) (Awasthi et al., 2021)

## Roadmap to establishing a tissue culturebased gene transfer system

#### Regeneration

- Which cultivars will regenerate?
- Media composition
  - ► Hormones
  - Sugars
  - Macronutrients
- Starting plant tissue type
- Lighting conditions





#### Transformation

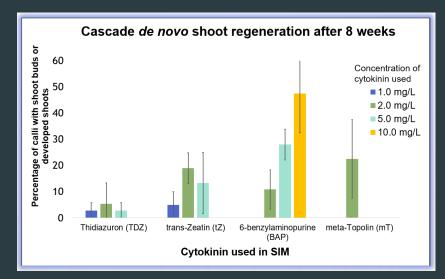
- Which strains of Agrobacterium to use
- How much Agrobacterium inoculum to use
- Which visual and selectable marker genes to use
- Starting plant tissue type
- Techniques to help Agrobacterium deliver DNA to more cells

## Experiments testing regeneration

#### Screened for shoot regeneration capacity in several public hop cultivars



#### Optimization for media hormone content in individual cultivars



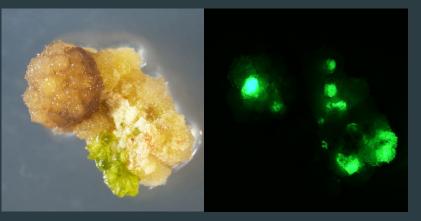


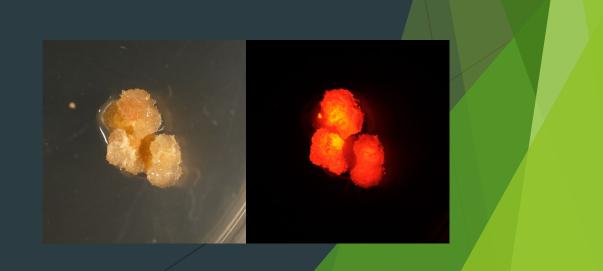
## Experiments testing transformation

- Using visual marker gene expression as a basis to see how well gene delivery worked
- > Agrobacterium strains compared
- Several methods to try to increase gene delivery from Agrobacterium
- Compared utility of different marker genes

50 Percentage of explants with GFP 45 40 expressing sector 35 30 25 20 15 10 5 LBA 0m LBA 5m LBA 10m AGL1\_0m AGL1 5m AGL1 10m Treatment

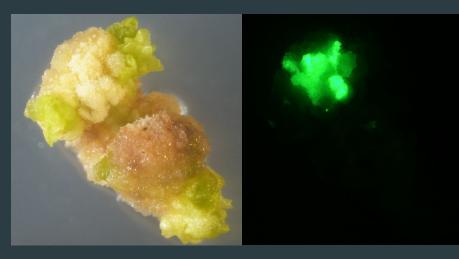






## Status of our in vitro transformation system

- Working primarily with Cascade and Fuggle
- ► Gene delivery to stem tissue is low
- Need to optimize selection to balance regeneration and transformed tissue
- Testing more methods to boost transformation and further improve regeneration efficiency

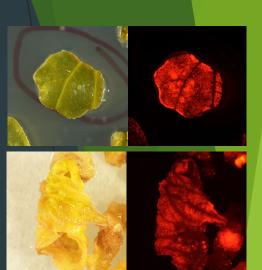




## Some promising directions to explore

Plant tissues other than stem might be better-suited for transformation





An alternative approach using "hairy roots"



- Development ("DEV") genes to facilitate transformation
- In planta transformation methods still a very new technology, but would allow us to bypass tissue culture

# What traits could be addressed with genetic engineering/editing?

#### Traits benefitting growers:

- Disease resistance traits specific to common pathogens
- Plant height
- Flowering time—early or later-maturing varieties
- Traits benefitting brewers/end consumers:
- Storage stability
- Altered bittering and aroma qualities
- Medicinal compound production?



Prerequisites for applying gene editing to improve agricultural traits

A viable transformation system in the cultivar we want to improve

Genome sequencing data



 $\blacktriangleright$  Genes  $\rightarrow$  Traits information - genetics research

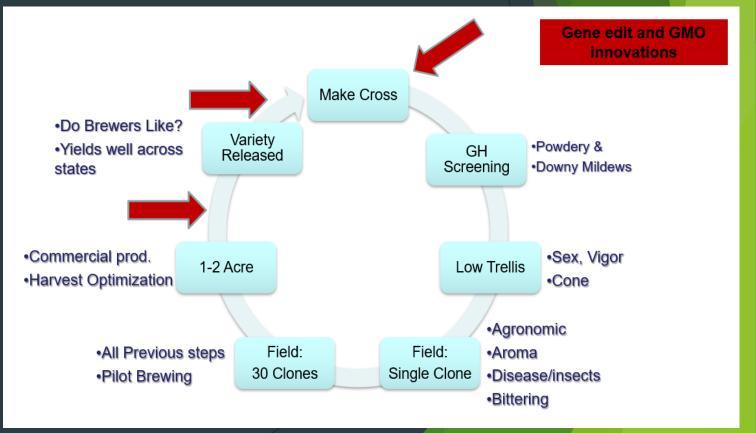


- Transformation techniques are the bottleneck for implementing gene editing in hop
- It is challenging, but the potential benefits are worth it
- Need to have more gene functional data for gene editing to be of most benefit
  - CRISPR can enable genetics research, helping to speed up discoveries

## Summary

GE is not something that can substitute for breeding, but it can:

- Complement it fill in gaps with respect to specific things that would be difficult to address through breeding
- Support it be used as a research tool to uncover gene functional information to inform breeders



## Acknowledgements



We thank USDA-NIFA for their support through AFRI grant #2021-67013-34739

Also, the awesome research team and invaluable assistance we've had on this project



Steve Strauss John Henning Dave Gent Pls



Michele Wiseman



Tanner Whiting (Undergraduate researcher)

Experimental work





Cathleen Ma Goralogia Strauss Lab

## R&D collaboration

We are seeking opportunities for partnership with industry groups to continue developing these tools in hop

Contact details for interest and/or additional info:

steve.strauss@oregonstate.edu

chris.willig@oregonstate.edu