

A hairy root-to-shoot transgene excision system for improved transformation and clean editing in clonally propagated plants

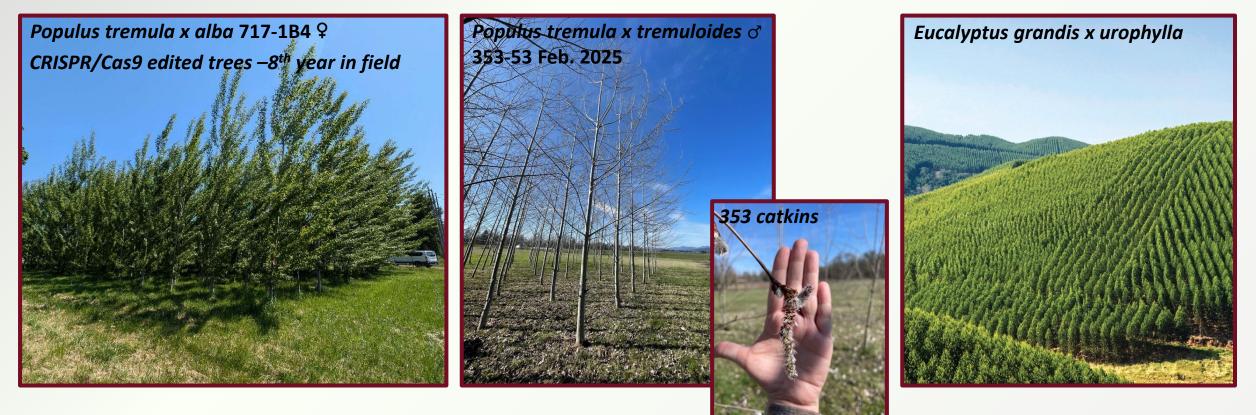
Greg Goralogia Laboratory of Steve Strauss

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College of Forestry Forest Ecosystems and Society



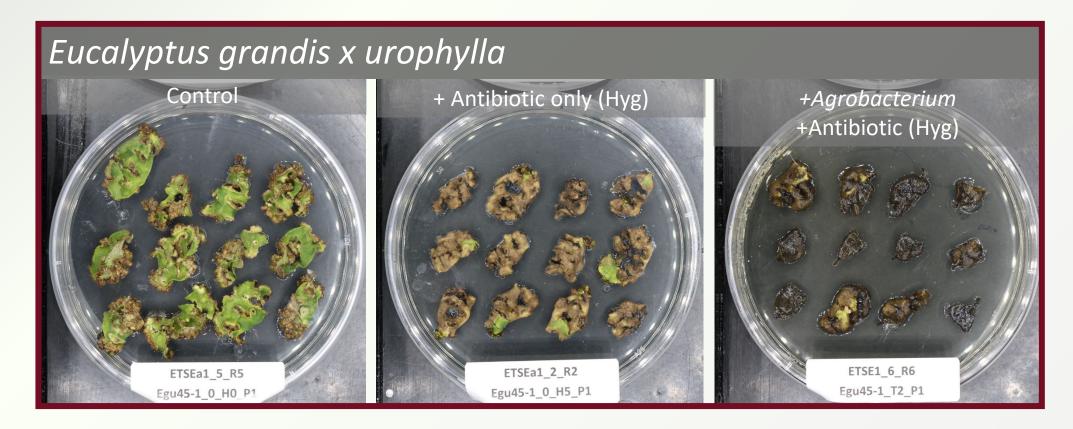
We work in tree species which have unique challenges for biotechnology



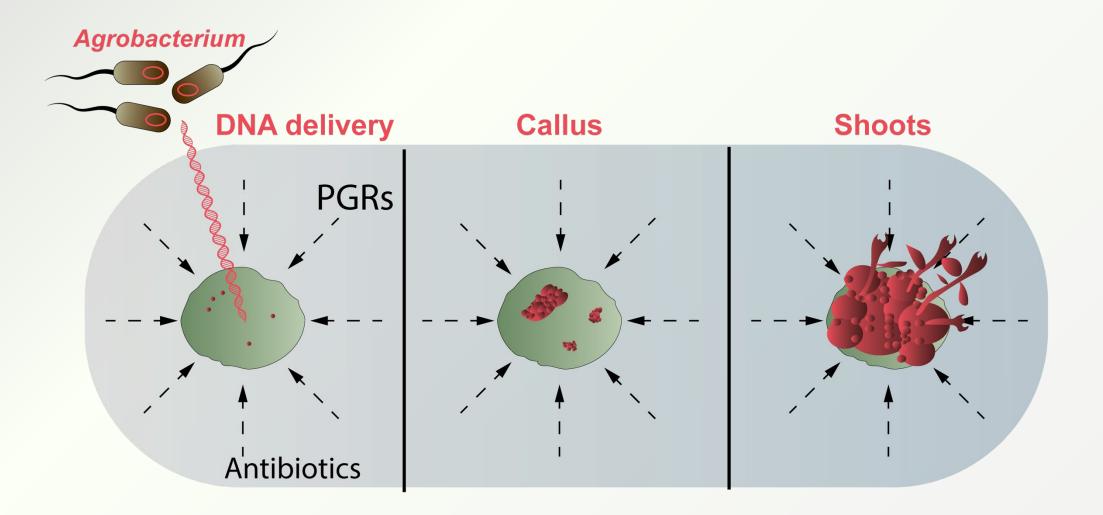
- Long testing cycles
- Regulatory challenges
- Clones are wide interspecific crosses, products of long breeding programs

Many tree species are difficult to transform

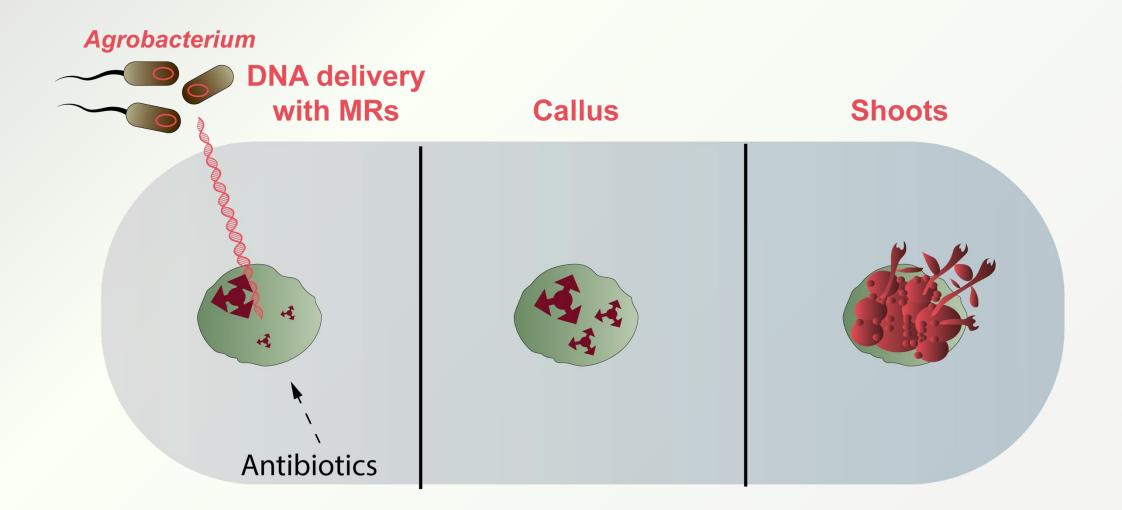
- Elite clones, not seed-derived
- High heterozygosity: each genotype a new adventure in vitro
- High physiological diversity and common defense responses



Conventional transformation methods rely on exogenous phytohormones supplied in the culture medium



Morphogenic regulators spur developmental reprogramming via delivere DNA –sometimes in the absence of exogenous PGRs



Types of MR genes we have studied in poplars or eucalypts – many both *in vitro* and *in planta*

- LEC 1, 2 LEAFY COTYLEDON
- EBB1 EARLY BUD BREAK 1 (ESR family)
- BBM BABY BOOM
- WOX 5, 11 -- WUSCHEL RELATED HOMEOBOX
- WUS WUSCHEL
- GRF-GIF GROWTH REGULATOR FACTOR 4 and GRF INTERACTING FACTOR 1
- Agrobacterium growth promoting genes
- rol Hairy root-inducing genes Agrobacterium

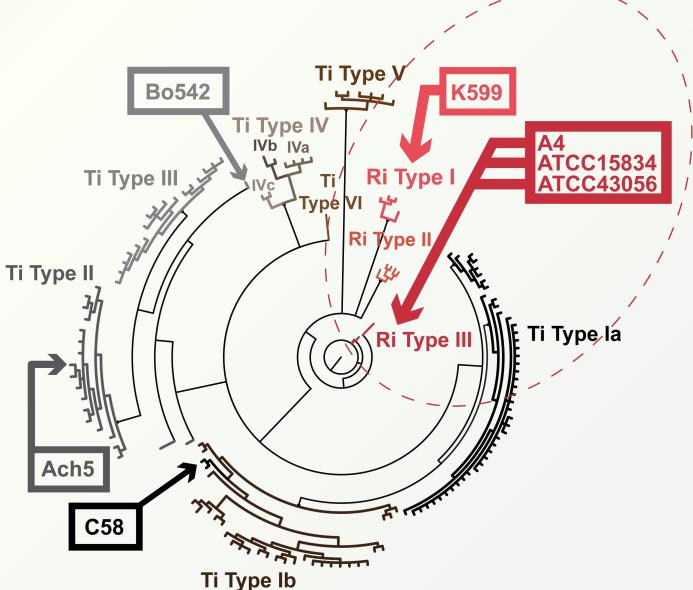
Most have failed with simple overexpression, or given highly genotype-specific enhancement or inhibition

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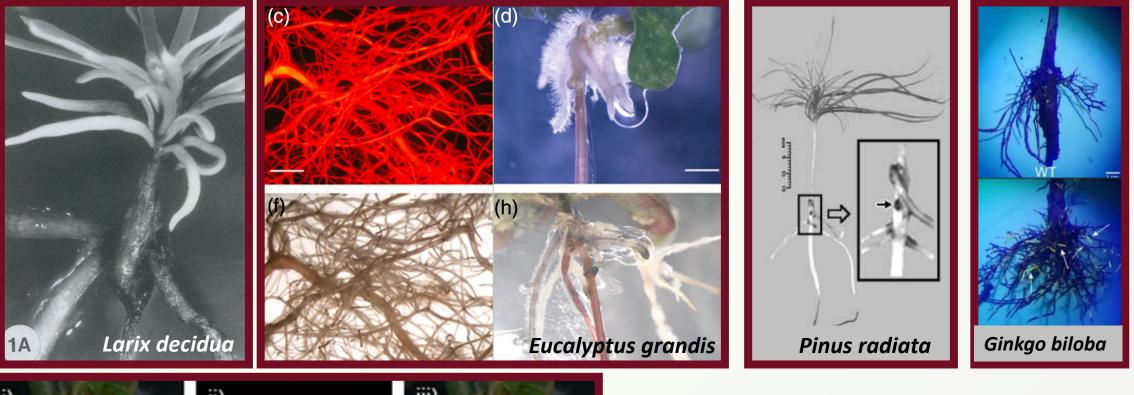
Hairy root disease is caused by unique T-DNA genes contained in Ri plasmids

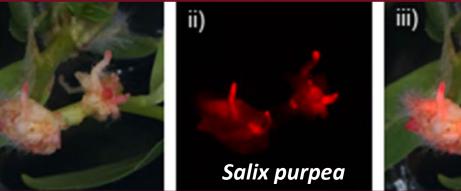


Hairy roots in hydroponically grown tomato



Hairy root *rol* genes are an effective way to generate transgenic tissues across diverse tree species





Huang et al. 1991 *IVCDB-Plant*, Placencia et al. 2016. *Plant Biotech J.*, Li et al. 2003 *EJ Biotech*, Gomes et al. 2019 *FIPS*, Du et al. 2025 *PNAS*.

Hairy root transformation is effective in many recalcitrant clonal specialty crops



Vitis vinifera cv. Syrah

Hairy root transformation is becoming popular for genotype- independent generation of "mostly" normal looking transgenic plants

Plant Biotechnology Journal	000 8 0 0
Plant Biotechnology Journal (2023), pp. 1–3	doi: 10.1111/pbi.14096
Brief Communication <i>Rhizobium rhizogenes</i> -mediated hairy root in plant regeneration for bioengineering citrus	duction and
Manikandan Ramasamy ¹ (), Michelle M. Dominguez ¹ , Sonia Irigoyen ¹ (), Carmen S. Pad Kranthi K. Mandadi ^{1,2,3,} * ()	illa ¹ and
 ¹ Texas A&M AgriLife Research & Extension Center, Weslaco, TX, USA ² Department of Plant Pathology and Microbiology, Texas A&M University, College Station, TX, USA ³ Institute for Advancing Health Through Agriculture, Texas A&M AgriLife, College Station, TX, USA 	
	Rooted transgenic plants

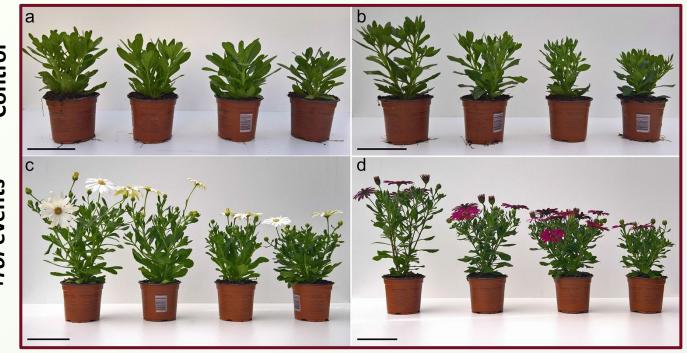
~4-6 weeks

~10-12 weeks

~2 weeks

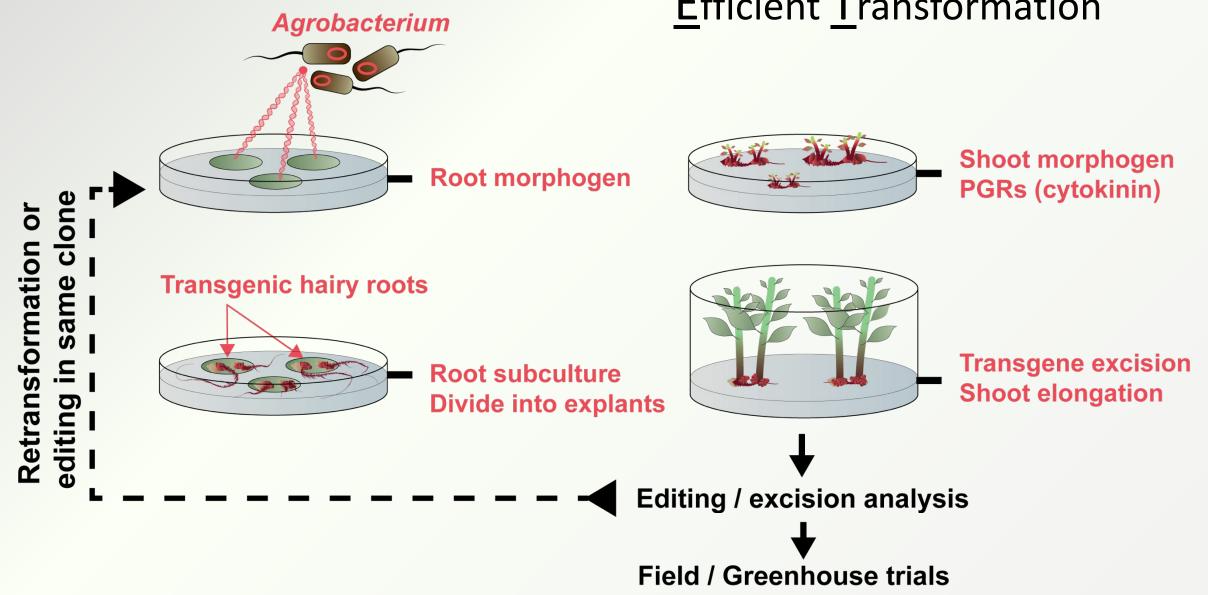
+*rol* events Contro

Osteospermum fruticosum (cape daisy)



rol transgenics often show dwarfism and changed floral timing and architecture

A concept for <u>Root Excision System</u> for <u>Efficient Transformation</u>



Root to shoot regeneration is a synthesis of older ideas

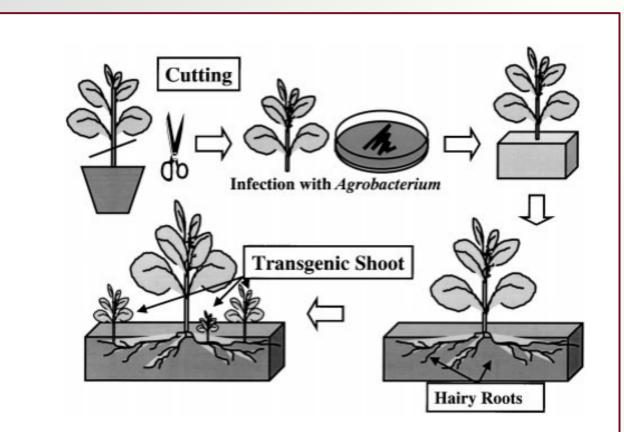
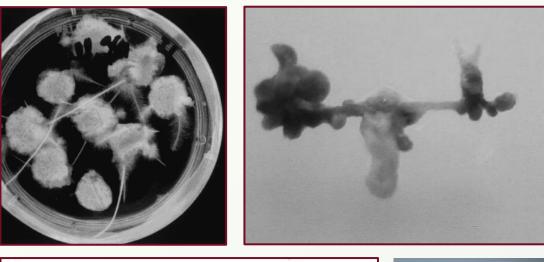
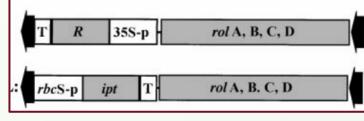


FIG. 8. Outline of *in vivo* transformation using cuttings. Cuttings are infected with *Agrobacterium* containing the *rol*-type MAT vector. After their rooting, marker-free transgenic shoots are induced from hairy roots by the light.





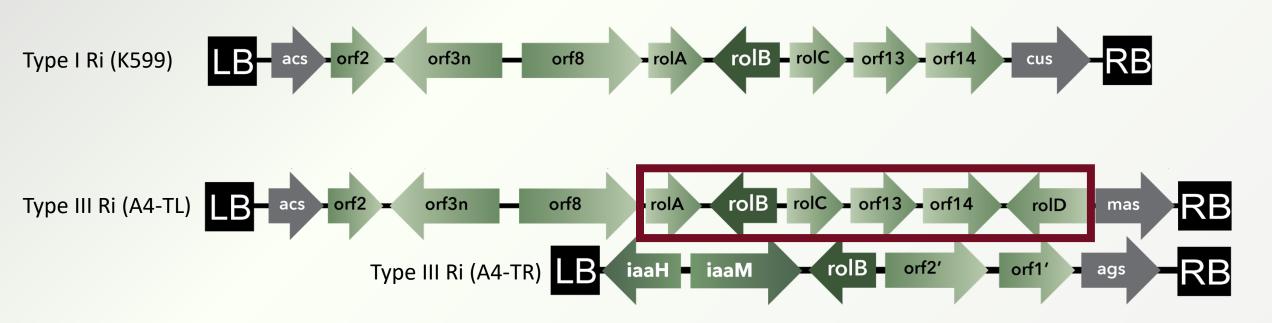
from Ebinuma and Komamine, 2001 In vitro cell and developmental biology -Plant



Hiroyasu Ebinuma (Shinsu U. em, Nippon Paper Co.)

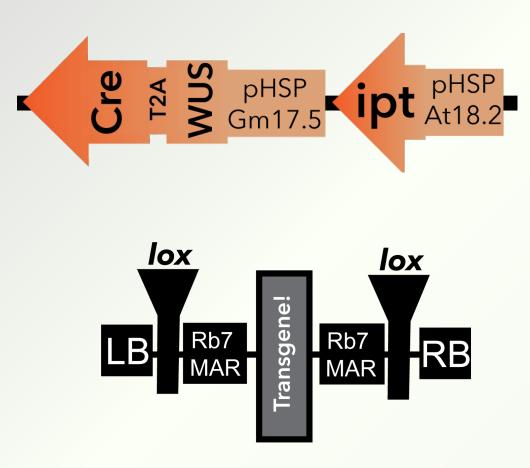
Let's go shopping for parts!

We selected a set of six *rol* genes from strain A4, a Type III Ri plasmid which is known to work in tree species



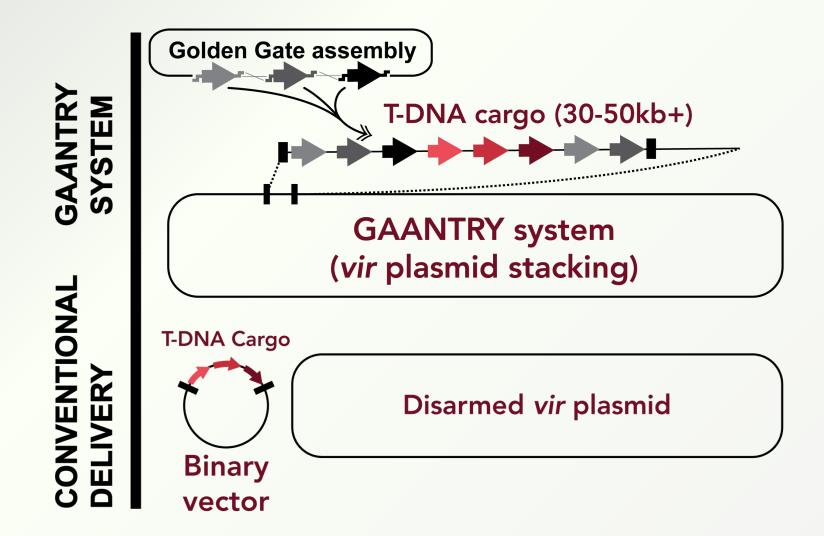
- rolB is required for hairy root formation
- Others including rolA, B, C, D, orf13 and orf14 quantitatively increase hairy root formation in many species

We selected **WUSCHEL** and **ipt** as shoot morphogens, **Cre-lox** for excision and chose heat shock as our induction system

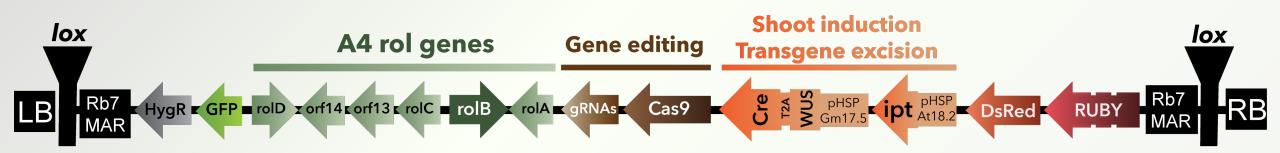


- *WUS* shown to induce shoot trans-differentiation from roots
- *ipt,* an *Agrobacterium* T-DNA gene which produces cytokinins and works well in our experimental system
- Heat shock induction is leaky, but we have tested all promoters in poplar
- Cre recombinase used to induce transgene excision, construct flanked by lox sites
- Rb7 MAR elements known to reduce DNA –methylation dependent transgene silencing

Assembly would be next to impossible without GAANTRY

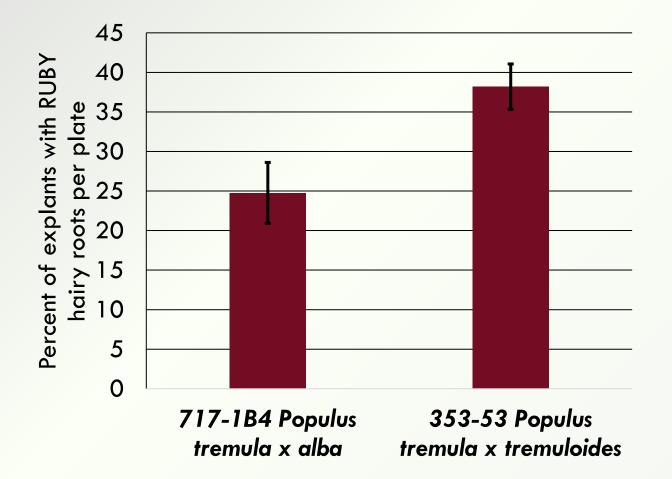


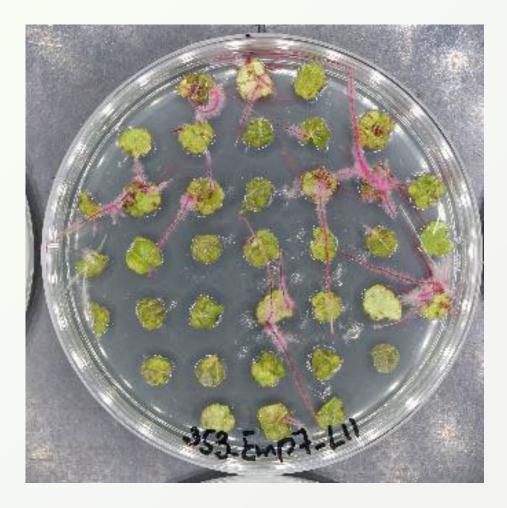
The "Kitchen sink" RESET construct



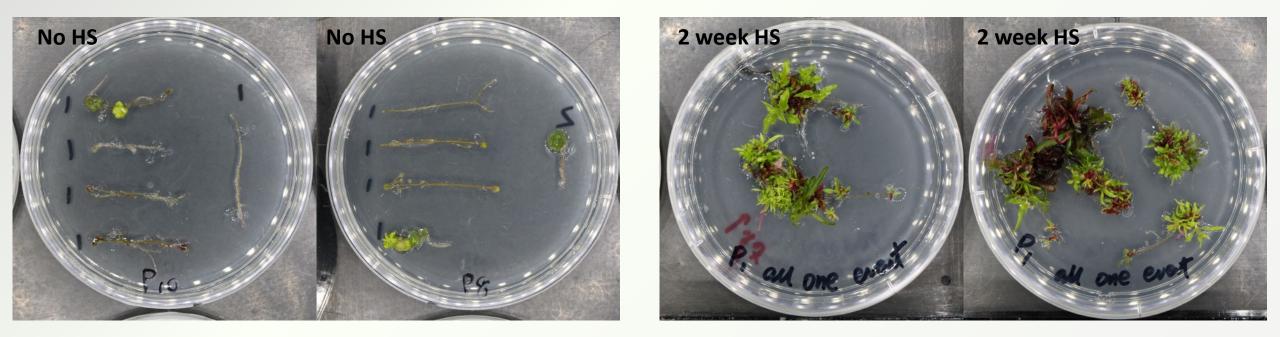
- 39kbp in size in this configuration
- 14 transcriptional units
- 16 independent peptides
- Included 3 marker genes to identify transgene insertion (GFP, DsRed, and RUBY)
- Includes gene editing through CRISPR-Cas9 (tRNA-arrays targeting *RGA1* gene)
- Hygromycin selection gene present but no selection was used in these experiments

We efficiently generated transgenic hairy roots in two poplar genotypes for regeneration analysis



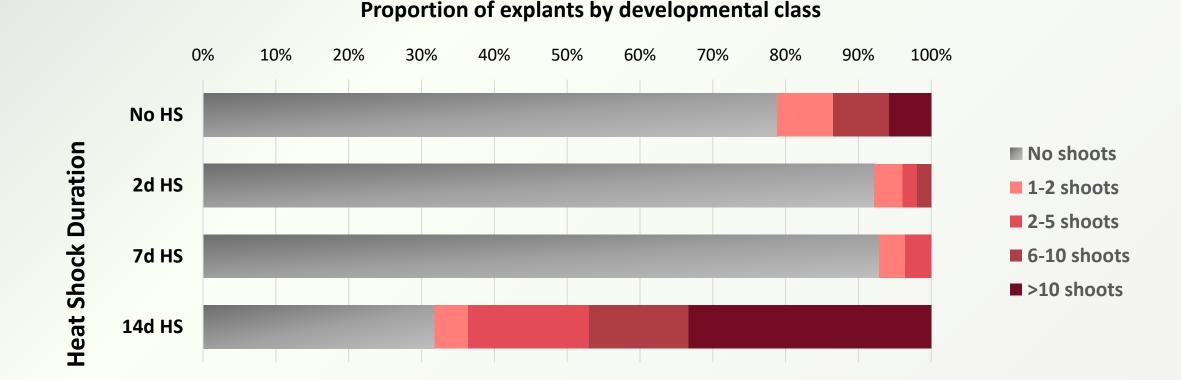


Two weeks of pulsed heat shock resulted in efficient shoot regeneration and excision from hairy roots



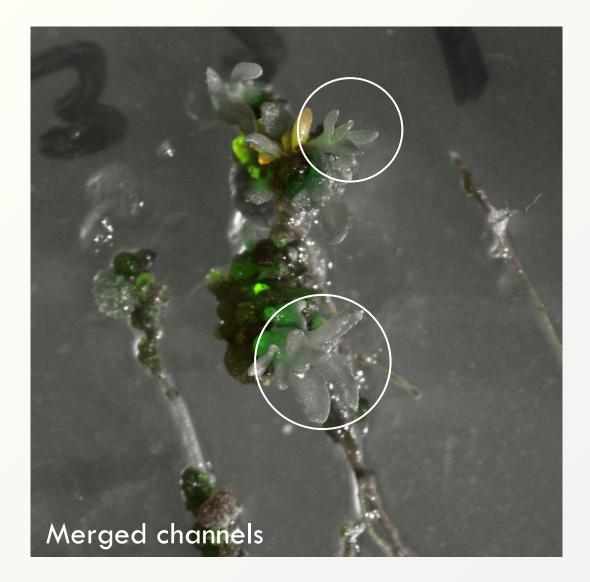
- Four hours heat shock at 39 degrees Celsius, different treatment durations
- RUBY vs. non-ruby shoots can be easily identified for propagation, then other reporters can be closer examined by fluorescent microscopy after isolation

Longer heat-shock (HS) duration improved shoot regeneration rates per explant (segments of hairy roots)



Using multiple reporters (GFP, DsRed and RUBY), we can find shoots with excised transgenes





After transfer of regenerating root explants, elongated shoots without marker genes can be identified

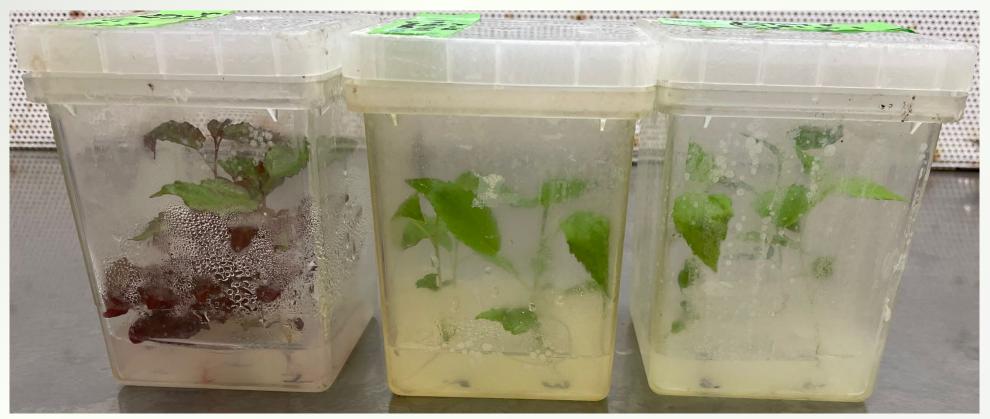


Mixtures of RUBY and green RESET shoots in late-stage propagation



Populations of putative excised RESET shoots for further molecular analysis

RESET excised shoots had normal phenotypes with continued *in vitro* culture and propagation



Unexcised transgenic

Putative excised event F

Putative excised event

We performed new transformations with the goal of estimating frequency and integrity of excision more reliably





We tracked individual hairy root insertion events through the transformation and excision process

Cre excision methods have been a constant frustration in this and other related projects



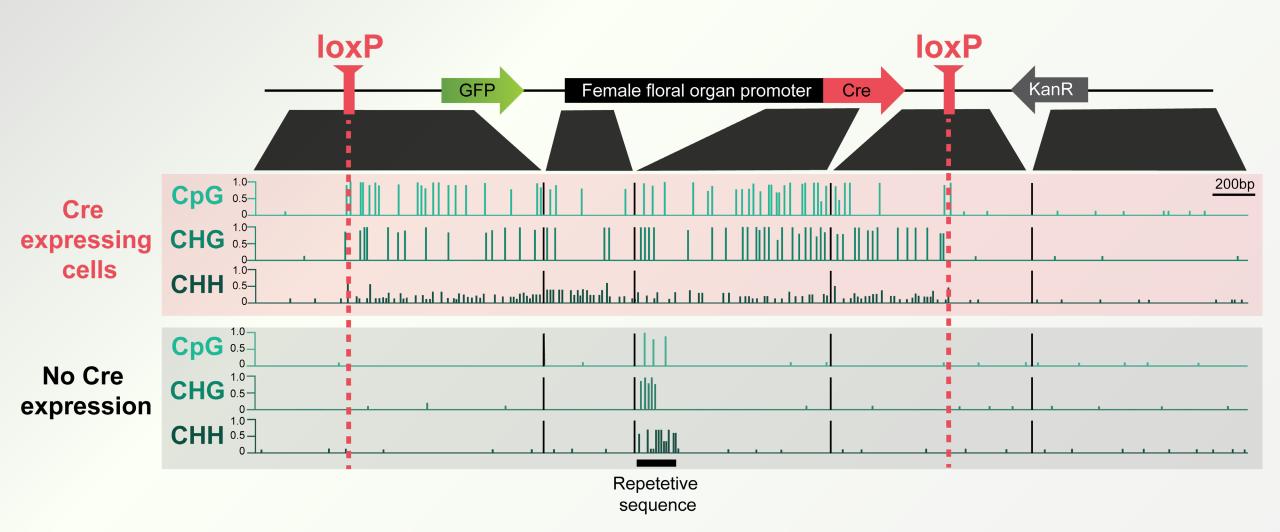


DNA methylation occurring in Cre-expressing cells inhibits loxP recombination and silences loxP-sandwiched genes

Ruochen Liu 💿, Qin Long 💿, Xiuping Zou 💿, You Wang and Yan Pei 💿

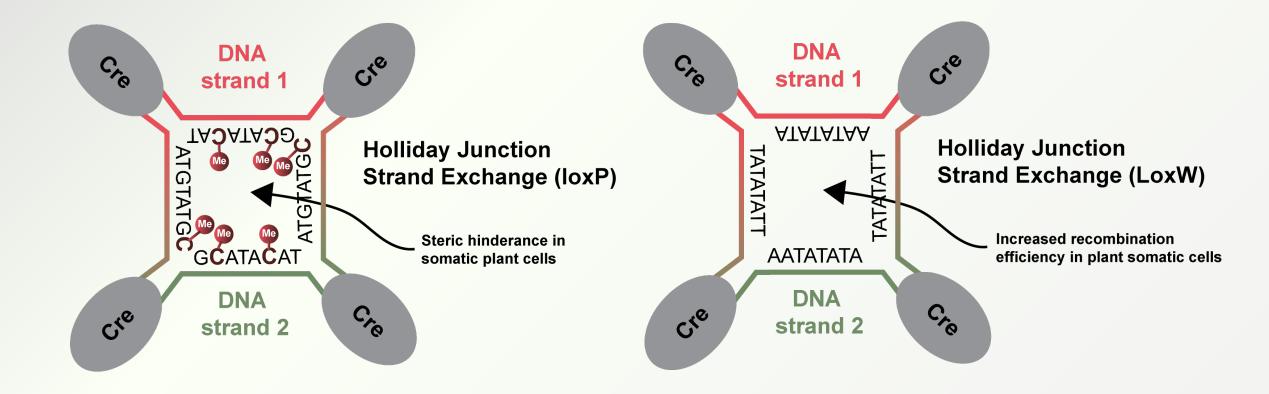
Chongqing Key Laboratory of Application and Safety Control of Genetically Modified Crops; Biotechnology Research Center, Southwest University, No. 2 Tiansheng Road Beibei, Chongqing 400715, China

Cre can methylate transgene regions flanked by its recognition site loxP

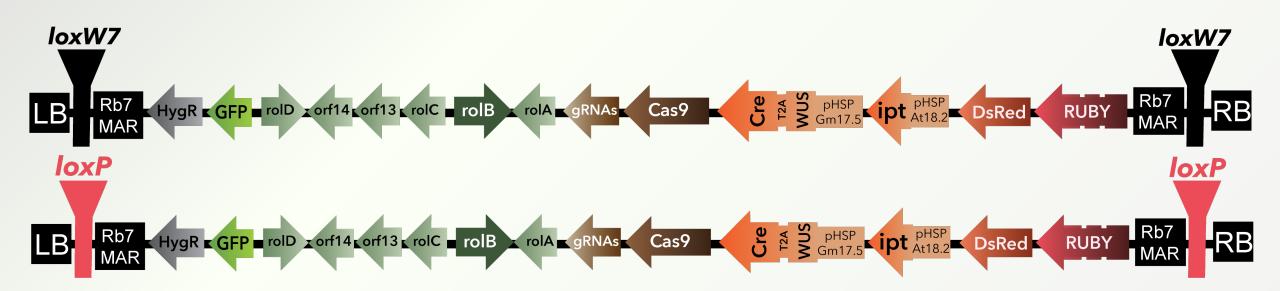


Modified from Liu et al. 2021 New Phytologist

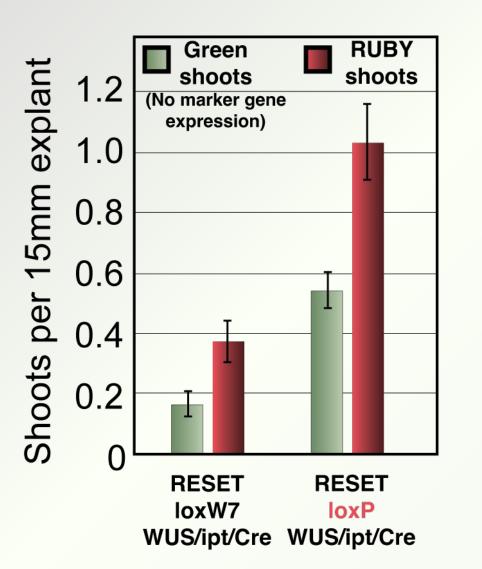
We made several variations of RESET constructs, including changes to flanking lox sites designed to be resistant to DNA methylation



We compared rates of excision of the two lox sites

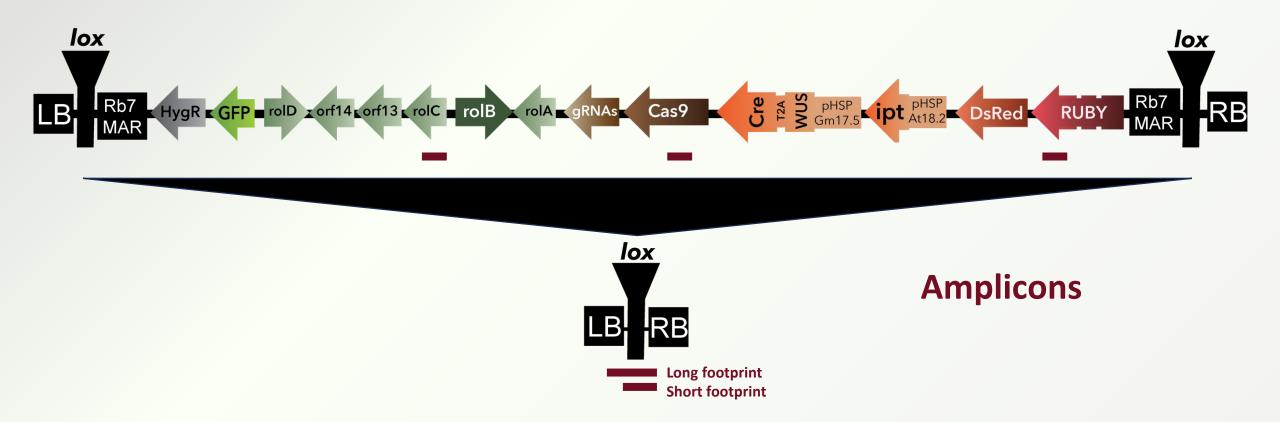


loxP-flanked RESET constructs gave the highest shoot regeneration after heat shock induction of transgenic roots



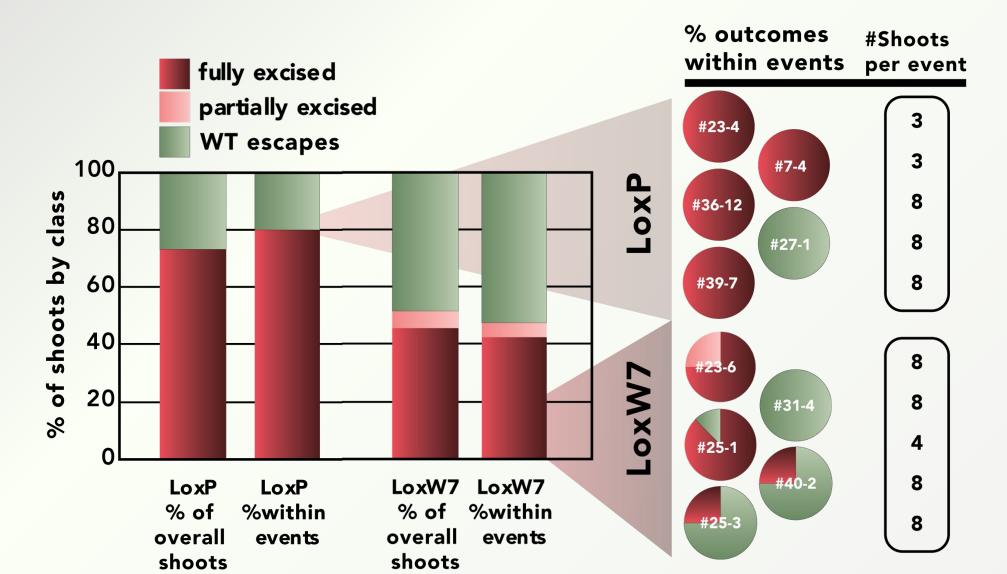
- 3.3x higher green shoot induction in loxP vs. loxW7
- 2.8x higher ruby shoot induction in loxP vs. loxW7
- Hairy root explant position (root tip, middle sections, or end) was not significant factor in regeneration outcome
- We kept multiple shoots originating from the same "mother" hairy root insertion event for detailed molecular analysis

To figure out if the shoots without RUBY, GFP, and DsRed were completely excised, we used a panel of PCR amplicons



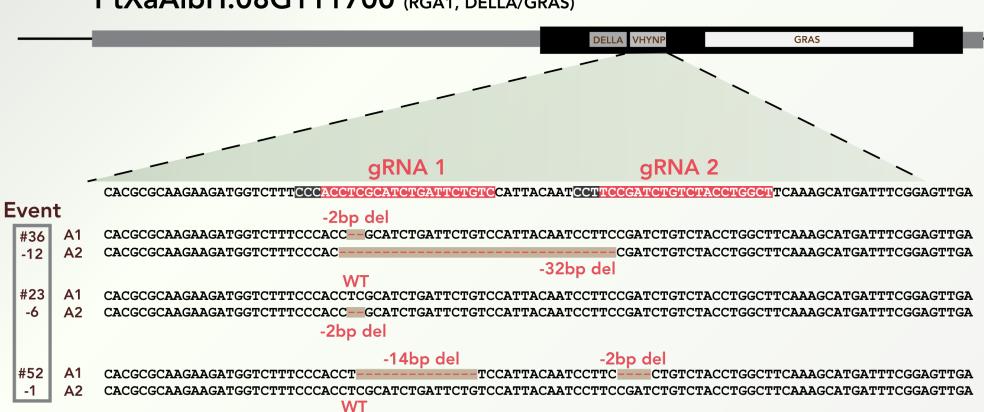
- The presence of any transgene band eliminated a shoot as being categorized fully excised (partial chimera)
- Two footprint amplicons were included in case of truncation of the T-DNA left border

While many escape shoots were found, high rates of complete excision were found in product shoots



Are they edited?

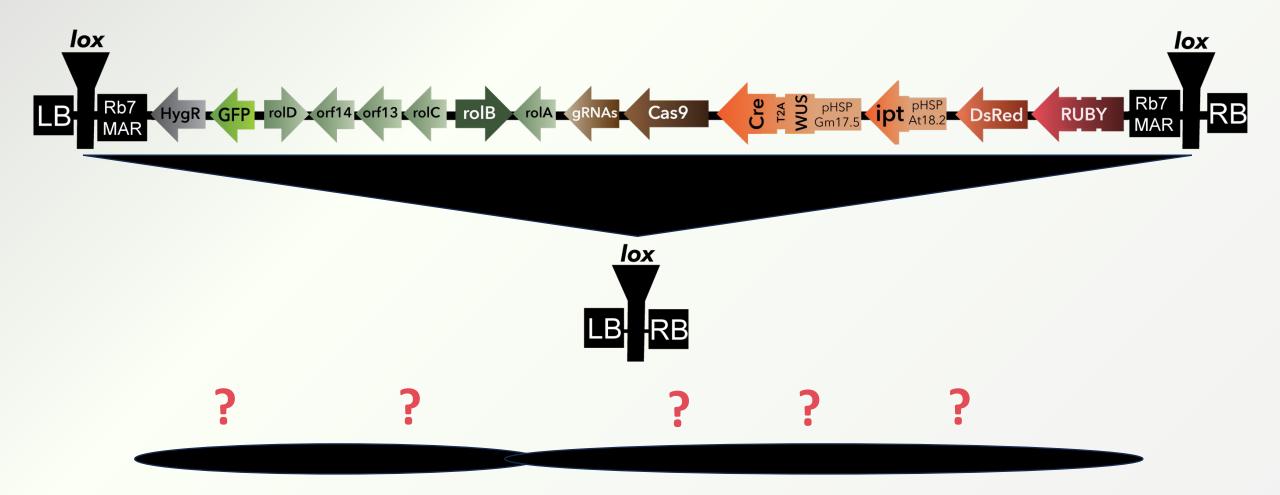
-We found high rates of editing among excised RESET shoots



PtXaAlbH.08G111700 (RGA1, DELLA/GRAS)

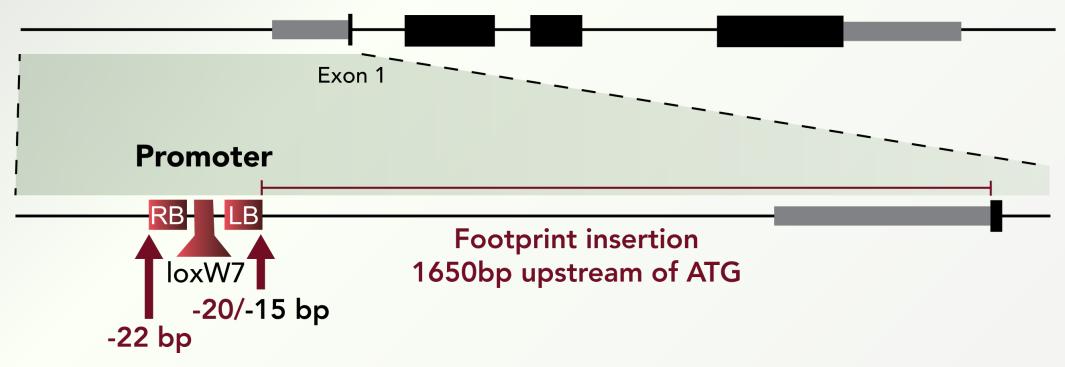
Analysis is ongoing but so far 6/10 events (60%) were edited to fixation in at least one allele

Where did our transgenes insert? How did the excision process resolve the footprints into final configuration?

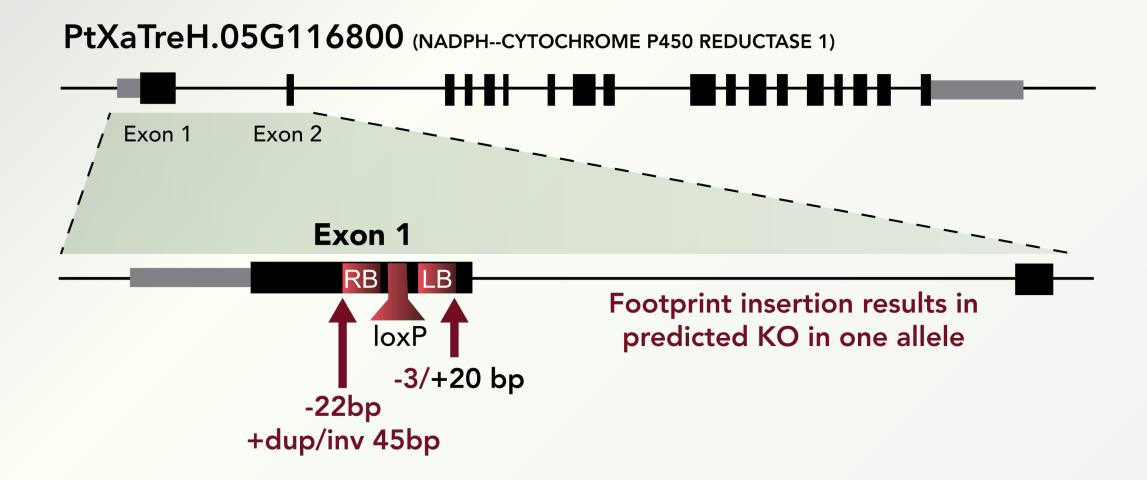


We used TAIL-PCR to find where transgenes landed and resolved into smaller footprints

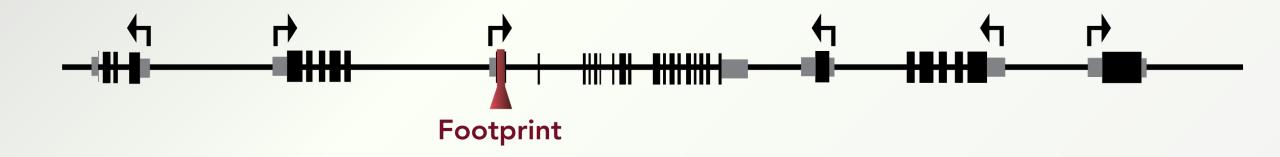
PtXaAlbH.06G051600 (PLATZ TRANSCRIPTION FACTOR)



We did find some events where the footprint inserted into an exon of one allele



Could these one day be considered clean edits for regulatory purposes? Are surrounding genes impacted?

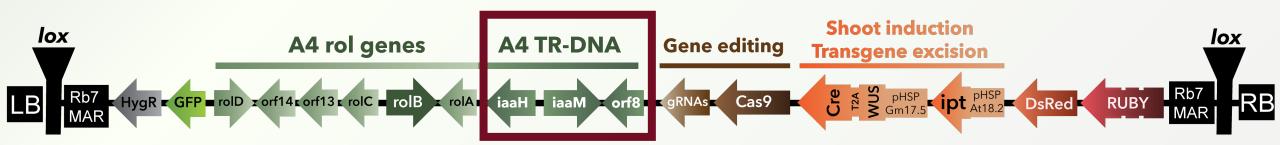


- Gene expression analysis of surrounding genes in characterized events to better understand local genetic impacts
- ddPCR to understand low abundance copy number variation of any residual transgenic cells
- Whole genome sequencing to confirm TAIL-PCR results and characterize excision resolution of footprints

Does this work in other, more recalcitrant species?



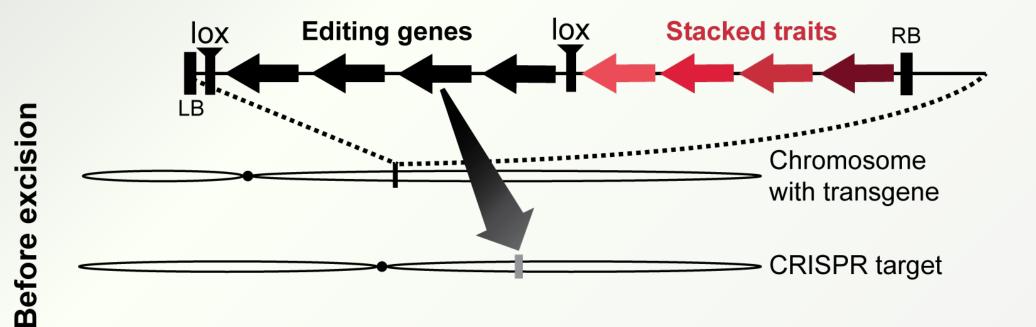
Eucalyptus grandis x urophylla RESET composite plants



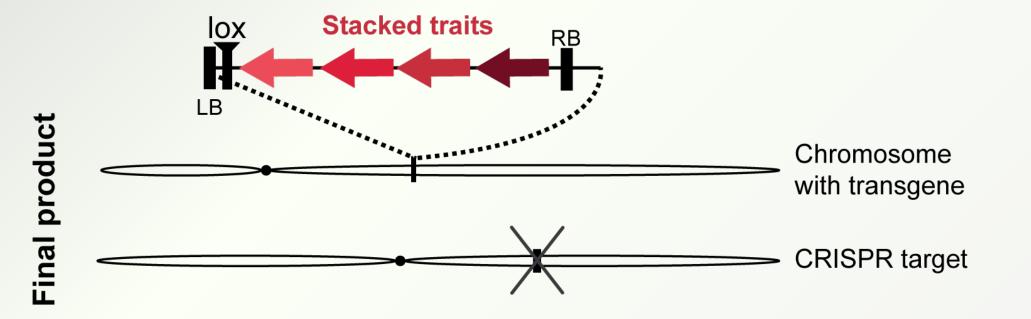
Hairy root to shoot methods are functional for editing and transformation in clonally propagated plants

- RESET system gave high rates of hairy root transformation, and especially with loxP-flanked borders gave efficient heat induced regeneration and transgene excision from root tissue
- High rates of editing are seen so far in excised shoots
- Significant numbers of escape shoots found which should be easily eliminated through adding selection during hairy root subculture
- Though complex, we hope this system will function broadly in clonal woody plant species

We envision this system being used simultaneous editing and transgene insertion, enabling synthetic biology approaches



....resulting in products with traits that could make downstream processes simpler, all in one operation



Acknowledgements: People





Victoria Conrad Undergraduate

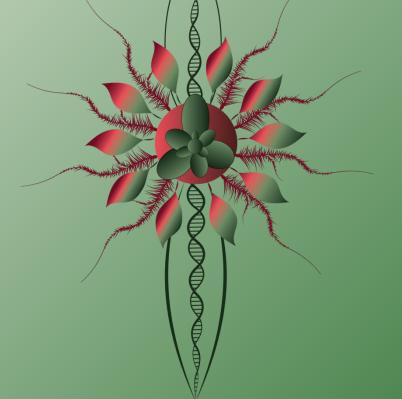
Sydney Gould Undergraduate

Cathleen Ma Tissue culture and transformation

Peremyslova Tissue culture and transformation

Kate

Steve Strauss Professor FES



Scientific assistance

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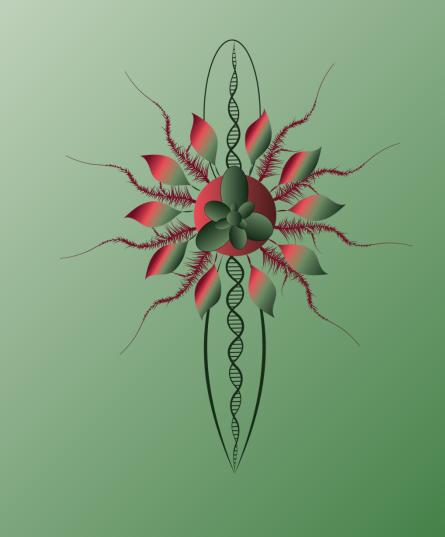
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Thanks for listening!